Eco. Env. & Cons. 29 (2) : 2023; pp. (915-920) Copyright@ EM International ISSN 0971–765X

DOI No.: http://doi.org/10.53550/EEC.2023.v29i02.062

Seaweeds- A Source of Potential Bioactive Compounds

Prasanthi Narra* and Annadurai D.

Centre of Advanced Study in Marine Biology, Annamalai University, Parangipettai, Tamilnadu, India

(Received 8 December, 2022; Accepted 13 February, 2023)

ABSTRACT

Seaweeds as a potential source of diet have been used in Japan, Korea and China since prehistoric times. In fact the usage of kelps dates back to 5th century in China. Seaweeds are having unique bioactive compounds and its composition, and nutritional benefits, its structures have been widely studied and are currently being therapeutically used for many medical conditions. Their sustainability in harsh environmental conditions is another unique characteristic that is of researcher's interest. In the current review, we have gathered important information on various bioactive compounds such as Complex Polysaccharides, Unsaturated Fatty Acids, Dietary Fibres, Polyphenolic compounds from seaweed, Phloroglucinol derivatives, Porphyrin derivatives, Proteins and Mineral constitutions, and other Seaweed Phytochemicals. Although are part of daily food in coastal regions of west, not much research is focused in Indian coastal areas to extract these bioactive components or to utilize these algal blooms. We need to utilize our regional algal blooms and proper techniques must be developed and streamlined for using our natural bioresource.

Key words: Algal blooms, Seaweeds, Bioresource, Bioactive compounds

Introduction

Since historic times, there has been a traditional use of seaweeds as food in China, Japan and the Republic of Korea [http://www.fao.org/3/y4765e/ y4765e0b.htm]. It has now gained more acceptances globally for its high nutritious value and being exotic component in menu. Literature suggests that around 21 species are used in everyday cuisines in Japan, six of them since the 8th century. Seaweeds accounted for more than 10% of the Japanese diet reaching out to 20% increase in its usage for the last 10 years. Seaweeds are currently pointed as the plant-origin sea greens from the future, earning already the status of "superfoods", which is a market term currently used due to consequence of their superior nutritional profile and richness in bioactive phytochemicals (Yuan et al., 2018; Pereira et al., 2012; Pirian et al., 2017). Seaweeds are now commercially important, renewable resource. Seaweeds are known to contain significant quantities of proteins, lipids, minerals and vitamins (Norziah et al., 2002; Sanchez-Machado et al., 2004; Van Ginneken et al., 2011). Seaweeds are very low in fat, high in mono and polyunsaturated fatty acids, and are very rich in carbohydrates, dietary fibers, proteins, containing all the essential amino acids, and vitamins such as A, B, C, E, those are usually absent in vegetables (MacArtain et al., 2008). In addition edible seaweeds are more enriched with several minerals, like sodium (Na), potassium (K), magnesium (Mg), phosphorous (P), iron (Fe), iodine (I), and zinc (Zn) (Rupérez et al., 2002; Bocanegra et al., 2009). Seaweeds also contain bioactive compounds in the form of polyphenols, carotenoids, vitamins, phycobilins, phycocyanins, and polysaccharides, among others those are known to benefit human health (Kadam *et al.*, 2010). Several authors report the consumption of seaweeds could contribute to higher intakes of K, that help to balance dietary Na/K where the higher levels of Na in seaweeds could be a therapeutic use for no salt replacement (Circuncisão *et al.*, 2018). In this review, we try gathering information and important literature on the bioactive molecules those make seaweeds unique and nutritious.

Protein

Since seaweeds are recognized as novel functional foods there is a need to characterize the composition of seaweeds (Harnedy et al., 2011; Greenwood et al., 1951). The protein content is recently noticed as it gives the emerging challenges to improve food security by identifying alternative and sustainable protein supplements. According to some studies, protein content ranges from 0.67% to 45.0% in red seaweeds; 5.02% to 19.66% in brown seaweeds; and from 3.42% to 29.80% in green seaweeds. On a gram-for-gram basis, seaweeds have protein and amino acid contents those are comparable to those of beef; however, seaweeds are consumed in much smaller quantities (Lourenço et al., 2002; EFSA Panel on Dietetic Products, 2010; Misurcova et al., 2014; Qasim, 1991). It is important to understand that the amino acid composition of proteins present in seaweed is critical to determine the value of proteins to the human diet, particularly in achieving an acceptable intake of essential amino acids. However, the digestibility of seaweed protein must be thoroughly studied as those proteins within the gastrointestinal tract will significantly affect the nutritional value of the protein as the protein-polysaccharide interactions is known to reduce digestion efficiency considerably. Thus in recent years research is being carried on the amino acid compositions of the seaweeds. Studies suggest seaweeds are rich sources of lysine an essential amino acid often present in limited quantities in plant protein sources such as corn, maize, soy, rice, and wheat (Galland-Irmouli et al., 1999; Cherry et al., 2019).

Complex Polysaccharides

Among marine resources, seaweeds are well known natural sources of polysaccharides. Sulfated polysaccharides are of the most common polysaccharides found in the cell walls of seaweeds. Seaweeds contain 2.97-71.4% complex polysaccharides such as alginate, fucoidan, and laminarin in brown seaweeds; xylan and sulphated galactans, such as agar, carrageenan, and porphyran in red seaweeds; whilst ulvan and xylan are found in green seaweeds. Seaweed polysaccharides are atypical in structure when compared to terrestrial glycans. These are known to resist gastric acidity, horde digestive enzymes, and gastrointestinal absorption. Seaweed glycans therefore serve as fermentation substrates for specific gut microbial flora or facilitate substrate cross-feeding such as oligosaccharides and metabolic cross-feeding of SCFAs to root indirect proliferation of specific bacteria's (Rose et al., 2009; Timm et al., 2010; Belenguer et al., 2006; Macfarlane et al., 2012; Ríos-Covián et al., 2016). Alginate, an algal polysaccharide is widely used for various purposes in the food industry such as a stabilizer, thickening or emulsifying agent. As it is an indigestible polysaccharide, alginate may also be considered as a source of dietary fiber. Till date the bioactivities reported in the literature from red seaweeds such as Palmariapalmata and Porphyraspp and from brown seaweeds such as Undariapinnatifida and were associated with antihypertensive, antioxidant, and antidiabetic effects. Other published works has suggested that dietary fibers may protect against a number of cardiovascular and gastrointestinal diseases. Therefore these complex polysaccharides from seaweeds hold promise as a dietary source, providing their bioactivity is validated in humans (Goñi *et al.*, 2002; Urbano *et al.*, 2002).

Polyphenols

Seaweeds are a rich source of polyphenols, such as catechins, flavonols, and phlorotannins. Red and green seaweeds are also a source of bromophenols, phenolic acids, and flavonoids. Phlorotannins are the most abundant polyphenol found in brown seaweeds. Most polyphenols of plant originmust undergo intestinal biotransformation by endogenous enzymes and the gut microbiota prior to absorption across enterocytes and are converted to glycones and aglycones by endogenous β -glucosidases in the small intestine. The purported bioactivities of seaweed polyphenols include potential anticancer, antimicrobial and antioxidant activities. Few studies have reported inhibition of digestive enzymes, which may prevent lipid absorption and help maintain glucose homeostasis (Murugan et al., 2015; Wan-Loy *et al.*, 2016; Fernando *et al.*, 2016; Yang *et al.*, 2010; Farasat *et al.*, 2014; Machu *et al.*, 2015; Murray *et al.*, 2018).

Recent studies report that seaweed extracts containing high polyphenol content that could reduce fasting blood glucose, total cholesterol, and LDL-C in humans, but the few interventions conducted in humans have reported inconsistent findings for the effect of seaweed polyphenols on other biomarkers associated with risk of type 2 diabetes and cardiovascular disease, including postprandial blood glucose, fasting insulin, HDL-C, and triglycerides (Lee *et al.*, 2012; Ko *et al.*, 2013; Murray *et al.*, 2018; EFSA Panel on Dietetic Products, 2010, 2011; Gall *et al.*, 2015; Yoon *et al.*, 2017; Li *et al.*, 2017; Cherry *et al.*, 2019).

Terpenoids

In seaweeds, the main carotenoid and tetrapenoid compounds with potential application in the food industry is fucoxanthin and is widely used as preservative for prevention of lipid peroxidation in meat. There have been a lot of studies performed using Fucoxanthin *in vivo*. Results have shown to reduce the risk of diabetes and obesity, and accumulation of lipid in the liver; to decrease insulin resistance; and to improve the plasma lipid profile (Bedoux et al., 2006; El Gamal, 2010). Thus consuming seaweeds have potential effects on anti-diabetic, antiobesogenic and antioxidant bioactivities. Seaweeds also contain carotenoids present in seaweeds, such as lutein, β -carotene, zeaxanthin, echinenone, violaxanthin, and neoxanthin, investigation for their potential antiobesogenic, antidiabetic, or antioxidant bioactivities.

Dietary Fibres

Dietary fibre is an important factor for our digestive health and regular bowel movements. These dietary fibres also helps you feel fuller, can improve cholesterol and blood sugar levels and thus indirectly assist preventing diabetes, heart disease, bowel cancer and improving satiety, interactions with gastrointestinal hormones, suppressing post-prandial lipaemia, reduced glycaemic and insulin levels, as well as reduction of low grade inflammation. Many seaweed species contain similar or higher total fibre content compared with their terrestrial plant sources (Woerle *et al.*, 2006; UKPDS, 1998; Pan *et al.*, 1997). Our daily intake of food is failing to meet daily requirements for dietary fiber intake. The potential functional properties of dietary fiber are directly related with the viscous and water-binding properties of fibers present in the gastrointestinal tract. Dietary fiber components improve health through fermentation process by the colonic microbiota, which can enhance alterations in gut microbial composition and increase the production of health-associated volatile fatty acids such as acetate, propionate, and butyrates. These fiber-induced alterations to the microbiota composition and the associated metabolites produced are naturally altered increasingly those relate with the promotion of gastrointestinal, cardiometabolic, immune, bone, and mental health (EFSA Panel, 2012, 2017; Scientific Advisory Committee on Nutrition, 2015; Clark et al., 2013; Gibson et al., 2017).

Unsaturated Fatty Acids

Unsaturated fats are well known as 'healthy' fats and they're important to include as part of a healthy diet. These fats reduce the risk of high blood cholesterol levels and have other many health benefits when they are replaced with intake of saturated fats in the diet. Many studies have confirmed that Seaweeds are rich in unsaturated fatty acids and thus have been considered its utilization as nutraceutical product (Kendel *et al.*, 2015; Vessby *et al.*, 1980; Borkman *et al.*, 1991).

MUFA

Seaweeds are known for its utilization as nutraceutical due to its richness in nutritionally beneficial components compounds such as proteins, carbohydrates, antioxidants, minerals, dietary fibers, vitamins monosaturated fatty acids (MUFAs) and polyunsaturated fatty acids (PUFAs) (Garg *et al.*, 1992; Riserus *et al.*, 2009). MUFA is linked to changes in incretin responses and emptying gastric troubles, where in it elevates dietary MUFA the glucagon-like peptide (GLP-1) in both healthy and diabetic subjects which keeps the risk of T2DM low. In addition, it is also known to increase levels of adiponectin that reduces risk of T2DM (Dimopoulos *et al.*, 2006; Thomsen *et al.*, 2003).

PUFA

Since the healthy diets are already very rich in n-6 PUFAs, greater focus needs to be placed on incorporating n-3 PUFAs into the diet. The main challenge remains that dietary sources of n-3 PUFAs are readily available but in limited quantities. Also

when compared to MUFA, polyunsaturated fatty acids (PUFA) play an important role in biological functions both structural and physiological in nature (Esposito *et al.*, 2009).

Seaweed species encompass phyla those contain good source of dietary PUFA as their omega-6 and omega-3 fatty acids ratio ranged from 0.29 to 6.73. Foods rich in omega-6 PUFA have improved insulin sensitivity. Omega-3 PUFA also influences expression in lipid and carbohydrate metabolism (Esposito *et al.*, 2009).

Conclusion

Seaweed is rich source of nutrition containing both macro and micronutrients in abundant quantity than any land vegetables. It is an excellent source of macronutrients such as dietary fibres, protein, complex polysaccharides and micronutrients including B12, folate, calcium, magnesium, zinc, iron, and selenium. Also seaweed is a known to have great source of iodine. With all these excellent qualities, cultivation of seaweeds and regular intake of them is highly recommended. Not much research is done in including them under regular food diet. Also not much of them are quantified and utilized. In India there are rich algal blooms in south coastal zone, however not even 05% of the population utilize it under the diet unlike few countries like Japan. In this review we try gathering the most beneficial bioactive components of Seaweeds so the awareness is increased and may be its utilization under diet is introduced in India. Natural resource with abundant nutritional benefits must be considered to be cultivated and industries must work on developing the combinational diet supplements.

References

- Bedoux, G., Hardouin, K., Burlot, A.S. and Bourgougnon, N. 2014. Bioactive compounds from seaweeds: Cosmetic applications and future development. *Advances in Botanical Research*. 71: 345-379.
- Belenguer, A., Duncan, S.H., Calder, A.G., Holtrop, G., Louis, P., Lobley, G.E. and Flint, H.J. 2006. Two routes of metabolic cross-feeding between Bifidobacteriumadolescentis and butyrate-producing anaerobes from the human gut. *Applied and Environmental Microbiology*. 72(5): 3593-3599.
- Bocanegra, A., Bastida, S., Benedí, J., Ródenas, S. and Sánchez-Muniz, F.J. 2009. Characteristics and nutritional and cardiovascular-health properties of

seaweeds. Journal of Medicinal Food. 12(2): 236-258.

- Borkman, M., Campbell, L.V., Chisholm, D.J. and Storlien, L.H. 1991. Comparison of the effects on insulin sensitivity of high carbohydrate and high fat diets in normal subject. *Journal of Clinical Endocrinology and Metabolism.* 72(2) : 432–437.
- Cherry, P., O'Hara, C., Magee, P.J., McSorley, E.M. and Allsopp, P.J. 2019. Risks and benefits of consuming edible seaweeds. *Nutrition Reviews*. 77(5): 307-329.
- Circuncisão, A.R., Catarino, M.D., Cardoso, S.M. and Silva, A.M.S. 2018. Minerals from Macroalgae Origin: Health Benefits and Risks for Consumers. *Marine Drugs.* 16(11) : E400.
- Clark, M.J. and Slavin, J.L. 2013. The effect of fiber on satiety and food intake: a systematic review. *Journal of American College of Nutrition*. 32(3): 200–211.
- Diabetes Control and Complications Trial Research Group. 1993. The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. *New England Journal of Medicine*. 329(14) : 977-986.
- Dimopoulos, N., Watson, M., Sakamoto, K. and Hundal, H.S. 2006. Differential effects of palmitate and palmitoleate on insulin action and glucose utilization in rat L6 skeletal muscle cells. *Biochemical Journal*. 399(3): 473–481.
- EFSA Panel on Dietetic Products, Nutrition and Allergies. 2010. Scientific opinion on dietary reference values for carbohydrates and dietary fibre. *EFSA Journal*. 8(3): 1462.
- EFSA Panel on Dietetic Products, Nutrition and Allergies. 2010. Scientific opinion on the substantiation of health claims related to dietary fibre pursuant to Article 13(1) of Regulation (EC) No 1924/2006. EFSA Journal. 8(10): 1735.
- EFSA Panel on Dietetic Products, Nutrition and Allergies. 2011. Scientific opinion on the substantiation of health claims related to the replacement of mixtures of saturated fatty acids (SFAs) as present in foods or diets with mixtures of monounsaturated fatty acids (MUFAs) and/or mixtures of polyunsaturated fatty acids (PUFAs), and maintenance of normal blood LDL-cholesterol concentrations (ID 621, 1190, 1203, 2906, 2910, 3065) pursuant to Article 13(1) of Regulation (EC) No 1924/2006. EFSA Journal 9(4): 18.
- EFSA Panel on Dietetic Products, Nutrition and Allergies. 2017. Safety of Ecklonia cava phlorotannins as a novel food pursuant to Regulation (EC) No 258/97. *EFSA Journal*. 15: 5003.
- EFSA Panel on Dietetic Products. 2012. Nutrition and Allergies. Scientific opinion on dietary reference values for protein. *EFSA Journal*. 10(2): 2557.
- El Gamal, A.A. 2010. Biological importance of marine algae. *Saudi Pharmaceutical Journal*. 18(1): 1-25.
- Esposito, K., Maiorino, M.I., Ciotola, M., di Palo, C. P.,

Gicchino, M., Petrizzo, M., Saccomanno, F., Beneduce, F., Ceriello, A. and Giugliano, D. 2009. Effects of a Mediterranean-style diet on the need for anti-hyperglycemic drug therapy in partients with newly diagnosed type 2 diabetes: A randomized trial. *Annals of Internal Medicine*. 151(5): 306–314.

- Farasat, M., Khavari-Nejad, R.A., Nabavi, S.M.B. and Namjooyan, F. 2014. Antioxidant activity, total phenolics and flavonoid contents of some edible green seaweeds from northern coasts of the Persian Gulf. *Iranian Journal of Pharmaceutical Research*. 13: 163–170.
- Fernando, I.P.S., Kim, M., Son, K.T., Jeong, Y. and Jeon, Y.J. 2016. Antioxidant activity of marine algal polyphenolic compounds: a mechanistic approach. *Journal of Medicinal Food.* 19 (7): 615–628.
- Gall, E.A., Lelchat, F., Hupel, M., Jegou, C. and Pouvreau, V.S. 2015. Extraction and purification of phlorotanninsfrom brown algae In: Stengel DB, Connan S., eds. *Natural Products from Marine Algae* Methods and Protocols. 1st ed New York, NY: Springer Science+Business Media; 131–143.
- Galland-Irmouli, A.V., Fleurence, J., Lamghari, R., Luçon, M., Rouxel, C., Barbaroux, O., Bronowicki, J.P., Villaume, C. and Guéant, JL. 1999. Nutritional value of proteins from edible seaweed Palmariapalmata (dulse). *Journal of Nutritional Biochemistry*. 10(6): 353-359.
- Garg, A., Grundy, S.M. and Unger, R.H. 1992. Comparison of effects of high and low carbohydrate diets on plasma lipoproteins and insulin sensitivity in patients with mild NIDDM. *Diabetes*. 41(10): 1278– 1285.
- Gibson, G.R., Hutkins, R., Sanders, M.E., Prescott, SL., Reimer, R.A., Salminen, S.J., Scott, K, Stanton, S., Swanson, K.S., Cani, P.D., Verbeke, K. and Reid, G. 2017. Expert consensus document: the International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nature Reviews Gastroenterology* & Hepatology. 14(8): 491–502.
- Goñi, I., Gudiel-Urbano, M. and Saura-Calixto, F. 2002. In vitro determination of digestible and unavailable protein in edible seaweeds. *Journal of the Science of Food and Agriculture*. 82 (15): 1850–1854.
- Greenwood, D.A., Kraybill, H.R. and Schweigert, B.S. 1951. Amino acid composition of fresh and cooked beef cuts. *Journal of Biological Chemistry*. 193(1): 23-28.
- Harnedy, P.A. and FitzGerald, R.J. 2011. Bioactive proteins, peptides, and amino acids from macroalgae. *Journal of Phycology*. 47(2): 218-232.
- Kadam, S.U. and Prabhasankar, P. 2010. Marine foods as functional ingredients in bakery and pasta products. *Food Research International*. 4 (8): 1975-1980.
- Kendel, M., Wielgosz-Collin, G., Bertrand, S., Roussakis, C., Bourgougnon, N. and Bedoux, G. 2015. Lipid Composition, Fatty Acids and Sterols in the

Seaweeds Ulvaarmoricana, and Solieriachordalis from Brittany (France): An Analysis from Nutritional, Chemotaxonomic, and Antiproliferative Activity Perspectives. *Marine Drugs*. 13(9): 5606-5628.

- Ko, S.C., Lee, M., Lee, J.H., Lee, S.H., Lim, Y. and Jeon, Y.J. 2013. Dieckol, a phlorotannin isolated from a brown seaweed, Ecklonia cava, inhibits adipogenesis through AMP-activated protein kinase (AMPK) activation in 3T3-L1 preadipocytes. *Environmental Toxicology and Pharmacology*. 36(3): 1253–1260.
- Lee, D.H., Park, M.Y., Shim, B.J., Youn, H.J., Hwang, H.J., Shin, H.C. and Jeon, H.K. 2012. Effects of Ecklonia cava polyphenol in individuals with hypercholesterolemia: a pilot study. *Journal of Medicinal Food*. 15(11): 1038–1044.
- Li, Y.J., Fu, X., Duan, D., Liu, X., Xu, J. and Gao, X. 2017. Extraction and Identification of phlorotannins from the brown alga, Sargassumfusiforme (Harvey) Setchell. *Marine Drugs*. 15(2) : 49.
- Lourenço, S.O., Barbarino, E., De-Paula, J.C., Pereira, L.O. da, S. and Marquez, U.M.L. 2002. Amino acid composition, protein content and calculation of nitrogen-to-protein conversion factors for 19 tropical seaweeds. *Phycological Research*. 50(3): 233-241.
- MacArtain, P., Gill, C.I.R., Brooks, M., Campbell, R. and Rowland, I.R. 2008. Nutritional value of edible seaweeds. *Nutrition Reviews*. 65(12): 535-543.
- Macfarlane, G.T. and Macfarlane, S. 2012. Bacteria, colonic fermentation, and gastrointestinal health. *Journal of AOAC International*. 95(1): 50–60.
- Machu, L., Misurcova, L., Ambrozova, J.V., Orsavova, J., Mlcek, J., Sochor, J. and Jurikova, T. 2015. Phenolic content and antioxidant capacity in algal food products. *Molecules*. 20(1): 1118–1133.
- Misurcova, L., Bunka, F., VavraAmbrozova, J., Machù, L., Samek, D. and Kráèmar, S. 2014. Amino acid composition of algal products and its contribution to RDI. *Food Chemistry*. 151: 120-125.
- Murray, M., Dordevic, A.L., Bonham, M.P. and Ryan, L. 2018. Do marine algal polyphenols have antidiabetic, antihyperlipidemic or anti-inflammatory effects in humans? A systematic review. *Critical Reviews in Food Science and Nutrition*. 58(12): 2039–2054.
- Murray, M., Dordevic, A.L., Ryan, L. and Bonham, M.P. 2018. The impact of a single dose of a polyphenolrich seaweed extract on postprandial glycaemic control in healthy adults: a randomised cross-over trial. *Nutrients*. 10 (3): 270.
- Murugan, A.C., Karim, M.R., Yusoff, M.B.M., Tan, S.H., Asras, M.F.B.F. and Rashid, S.S. 2015. New insights into seaweed polyphenols on glucose homeostasis. *Pharmaceutical Biology*. 53(8): 1087–1097.
- Norziah, M.H. and Ching, C.Y. 2002. Nutritional composition of edible seaweeds *Gracilariachanggi*. Food Chemistry. 68(1): 69-76.
- Pan, X.R., Li, G.W., Hu, Y.H., Wang, J.X., Yang, W.Y., An,

Z.X., Hu, Z.X., Lin, J., Xiao, J.Z., Cao, H.B., Liu, P.A., Jiang, X.G., Jiang, Y.Y., Wang, J.P., Zheng, H., Zhang, H., Bennett, P.H. and Howard, B.V. 1997. Effects of diet and exercise in preventing NIDDM in people with impaired glucose tolerance. The Da Qing IGT and diabetes study. *Diabetes Care*. 20(4): 537–544.

- Pereira, H., Barreira, L., Figueiredo, F., Custódio, L., Vizetto-Duarte, C., Polo, C., Resek, V., Engelen, A. and Varela, J. 2012. Polyunsaturated fatty acids of marine macroalgae: Potential for nutritional and pharmaceutical applications. *Marine Drugs*. 10(9): 1920-1935.
- Pirian, K., Moein, S., Sohrabipour, J., Rabiei, R. and Blomster, J. 2017. Antidiabetic and antioxidant activities of brown and red macroalgae from the Persian Gulf. *Journal of Applied Phycology*. 29: 3151-3159.
- Qasim, R. 1991. Amino acid composition of some common Seaweeds. Pakistan Journal Pharmaceutical Sciences. 4(1): 49-54.
- Ríos-Covián, D., Ruas-Madiedo, P., Margolles, A., Gueimonde, M., de los Reyes-Gavilán, C.G. and Salazar, N. 2016. Intestinal short chain fatty acids and their link with diet and human health. *Frontiers in Microbiology*. 7: 185.
- Riserus, U., Willett, W.C. and Hu, F.B. 2009. Dietary fats and prevention of type 2 diabetes. *Progress in Lipid Research*. 48(1): 44–51.
- Rose, D.J., Keshavarzian, A., Patterson, J.A., Venkatachalam, M., Gillevet, P. and Hamaker, B.R. 2009. Starch-entrapped microspheres extend in vitro fecal fermentation, increase butyrate production, and influence microbiota pattern. *Molecular Nutrition and Food Research*. 53(S1): S121–S130.
- Rupérez, P. 2002. Mineral content of edible marine seaweeds. *Food Chemistry*. 79(1): 23-26.
- Sanchez-Machado, D.I., López-Cervantes, J., López-Hernandez, J. and Paseiro-Losada, P. 2004. Fatty acids, total lipid, protein and ash contents of processed edible seaweeds. *Food Chemistry*. 85(3): 439-444.
- Scientific Advisory Committee on Nutrition. 2015. Carbohydrates and Health. London, UK: TSO (The Stationery Office)
- Seaweeds used as Human food. FAO. Available at: http://www.fao.org/3/y4765e/y4765e0b.htm
- Thomsen, C., Storm, H., Holst, J.J. and Hermansen, K. 2003. Differential effects of saturated and monounsaturated fats on postprandial lipemia and glucagon-like peptide 1 responses in patients with type 2 diabetes. *American Journal of Clinical Nutrition*. 77(3): 605–611.
- Timm, D.A., Stewart, M.L., Hospattankar, A. and Slavin, J.L. 2010. Wheat dextrin, psyllium, and inulin produce distinct fermentation patterns, gas volumes, and short-chain fatty acid profiles in vitro. *Journal of*

Medicinal Food. 13(4): 961–966.

- United Kingdom Prospective Diabetes Study Group (UKPDS). 1998. Intensive blood-glucose control with sulfonylureas or insulin compared with conventional treatment and risk of complications in patients with Type 2 diabetes (UKPDS 33) *Lancet*. 352 (9131): 837-853.
- Urbano, M.G. and Goni, I. 2002. Bioavailability of nutrients in rats fed on edible seaweeds, Nori (*Porphyratenera*) and Wakame (*Undariapinnatifida*), as a source of dietary fibre. *Food Chemistry*. 76(3): 281–286.
- Van Ginneken, V.J.T., Helsper, J.P.F.G., de Visser, W., van Keulen, H. and Brandenburg, W.A. 2011. Polyunsaturated fatty acids in various macroalgal species from north Atlantic and tropical seas. *Lipids in Health and Disease*. 10: 1-8.
- Vessby, B., Gustafsson, I.B., Boberg, J., Karlstrom, B., Lithell, H. and Werner, I. 1980. Substituting polyunsaturated for saturated fat as a single change in a Swedish diet: Effects on serum lipoprotein metabolism and glucose tolerance in patients with hyperlipoproteinaemia. *European Journal of Clinical Investigation*. 10(3): 193–202.
- Vessby, B., Uusitupa, M., Hermansen, K., Riccardi, G., Rivellese, A.A., Tapsell, L.C., Nälsén, C., Berglund, L., Louheranta, A., Rasmussen, B.M., Calvert, G.D., Maffetone, A., Pedersen, E., Gustafsson, I.B., Storlien, L.H. and KANWU Study. 2001. Substituting dietary saturated for monounsaturated fat impairs insulin sensitivity in healthy men and women: The KANWU Study. *Diabetologia*. 44(3): 312–319.
- Wan-Loy, C. and Siew-Moi, P. 2016. Marine algae as a potential source for anti-obesity agents. *Marine Drugs*. 14: 222.
- Woerle, H.J., Szoke, E., Meyer, C., Dostou, J.M., Wittlin, S.D., Gosmanov, N.R., Welle, S.L. and Gerich, J.E. 2006. Mechanisms for abnormal post-prandial glucose metabolism in type 2 diabetes. *American Journal* of Physiology Endocrinology and Metabolism. 290(1): E67-E77.
- Yang, H.C., Zeng, M.Y., Dong, S.Y., Liu, Z. and Li, R. 2010. Anti-proliferative activity of phlorotannin extracts from brown algae *Laminaria japonica* Aresch. *Chinese Journal of Oceanology and Limnology*. 28: 122–130.
- Yoon, M., Kim, J.S., Um, M.Y., Yang, H., Kim, J., Kim, Y.T., Lee, C., Kim, S.B., Kwon, S. and Cho, S. 2017. Extraction optimization for phlorotannin recovery from the edible brown seaweed Ecklonia cava. *Journal of Aquatic Food Product Technology*. 26: 801–810.
- Yuan, Y., Zhang, J., Fan, J., Clark, J., Shen, P., Li, Y. and Zhang, C. 2018. Microwave assisted extraction of phenolic compounds from four economic brown macroalgae species and evaluation of their antioxidant activities and inhibitory effects on α-amylase, α-glucosidase, pancreatic lipase and tyrosinase. *Food Research International*. 113: 288-297.